

VERIFICATION OF TRANSLATION

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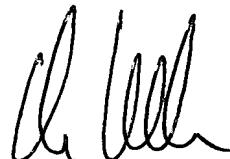
entitled: **ISOLATED AND PURIFIED HUMAN SOLUBLE
GUANYLYLCYCLASE $\alpha 1/\beta 1$ (hsGC $\alpha 1/\beta 1$)**

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am the translator of the documents attached and I state that the following is a true translation
to the best of my knowledge and belief of **German Patent Application: DE 198 37 015 filed
on August 14, 1998.**

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(Signature of translator)

**ENGLISH TRANSLATION OF DE 198 37 015****Isolated and purified human soluble guanylyl cyclase $\alpha 1/\beta 1$
(hsGC $\alpha 1/\beta 1$)**

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The technical field of the invention

The present invention relates to the expression of the cDNA clones for the subunits $\alpha 1$ (hsGC $\alpha 1$) and $\beta 1$ (hsGC $\beta 1$) of human soluble guanylyl cyclase and the subsequent purification of the active enzyme and the use thereof, the medical application of the expression of these clones by gene transfer, as well as antibodies to peptides derived from the sequence and the use thereof.

15 **Discussion of the state of the art**

The endogenous NO/cGMP signaling system mediates important functions such as vasodilation, inhibition of platelet aggregation, neurotransmission, and the immune response. In addition, it is involved in the development of various disease states such as ischemia-reperfusion and inflammatory injuries (Schmidt and Walter, 1994). Therefore, the NO/cGMP system has long been an important starting point for the development of novel drugs for the therapy of coronary heart disease, susceptibility to thrombosis, cardiac insufficiency, angina pectoris, cardiac-dependent pulmonary oedema, hypertensive crises, inflammatory states and cardiac infarction. Until now, such therapies have employed various so-called NO donors, e.g. nitroglycerin, that release NO, thereby replacing endogenous NO and activating soluble guanylyl cyclase (sGC) (Fig. 1). sGC forms cGMP, which mediates the effects of the NO/cGMP pathway via various intracellular receptor enzymes. The application of NO donors has two limitations: I) The repeated application of NO donors results in tolerance in the patient, i.e. loss of activity, when reapplied; II) NO reacts with O_2^- to form peroxynitrite, which is cytotoxic and less effective in activating sGC. Thus, direct activation of sGC by novel, non-NO-containing pharmaceuticals for therapy or gene transfer of sGC would be desirable strategies. Non-NO-releasing activators of sGC

are potentially lacking tolerance and NO toxication. Recently, "YC-1" (3-(5'-hydroxymethyl-2'-furyl)-1-benzylindazol) has been described as the first non-NO-releasing activator of sGC in platelets (Ko et al., 1994; Wu et al., 1995). YC-1 also activates purified sGC from bovine lung and potentiates the activation of NO (Friebe et al., 1996). The effect on human sGC has not been investigated. The human isoform of sGC is not yet available for pharmacological screening.

The NO receptor sGC consists of two subunits, α and β , which together form an enzymatically active heterodimer. Three different isoforms of the α and three of the β subunits have been described in the literature, although from different species. The best investigated isoforms, the bovine α_1/β_1 and the rat α_1/β_1 isoforms, have particular significance for cardiovascular research. Until recently, a human homologue of the bovine and rat sGC (e.g. sGC α_1 /sGC β_1 heterodimers) was unknown. cDNA sequences have been published that reportedly correspond to a human sGC α_3 and β_3 isoform (Giulii et al., 1992). While sGC β_3 showed a high homology to sGC β_1 (bovine/rat), the sGC α_3 sequence contained two restricted regions without homology to sGC α_1 (bovine/rat), here designated S1 and S2 (Fig. 2). In addition, it was unknown whether sGC α_3 and sGC β_3 can form a functional sGC heterodimer and which role the S1 and S2 regions play. Furthermore, no other human sGC subunit has yet been expressed as a protein. Recently, a sequence designated hsGC α_1 was published in the GeneBank (Accession No. U58855) that lacks the sequence differences to sGC α_1 from bovine and rat tissues in the regions S1 and S2. In addition, an alternatively spliced product of hsGC β_3 was recently published in the GeneBank (Accession No. AF020340) that was designated by the authors as an alternatively spliced form of hsGC β_1 . The physiological significance of this splice variant of hsGC $\beta_1/3$ is unclear. Thus, the question arises as to which of these isoforms is responsible for which physiological function in which cell types.

Furthermore, no antibodies to the human sGC α_1/β_1 are currently available that are monospecific, directed against the human sequences, or that have been shown to be suitable for immunoblots with human tissues. Peptide antibodies reported thus far only partially show these features: Harteneck et al. and Guthmann et al. used a peptide sequence (VYKVETVGDKYMTVSGLP) that is highly conserved in guanylyl

cyclases. Therefore, cross-reaction with particulate guanylyl cyclases (e.g. GC-C) can be expected. Guthmann et al. used a peptide sequence (YGPEVWEDIKKEA) identical to hsGC β 1 and a peptide sequence (KKDVEEANANFLGKASGID) identical to hsGC α 1 except for two amino acid exchanges. However, the function of these antibodies in immunoblots was only shown for *enriched* hsGC from human platelets. In addition, the antisera to hsGC α 1 detect a second, unspecific product. Humbert et al. and Koesling et al. used a peptide sequence (SRKNTGTEETE QDEN) of bovine sGC β 1 that partially (amino acids 1-10) overlaps with the peptide used here for the human sGC β 1 (amino acids 13-22) and is identical in this region. The C-terminus of this bovine peptide (amino acids 11-15), however, is clearly different from the human sequence. On the other hand, the antiserum to this peptide was not tested with a human protein but rather only used for immunoprecipitation of the bovine sGC.

Thus, for the human isoform α 1/ β 1 which is important for cardiovascular research, neither *native* protein nor *recombinant* protein, nor a one-step purification protocol, nor specific antibodies have been available. An approach for gene therapy (e.g. with adenoviruses or the like) has also not been described yet.

The technical problem of the invention

The NO-independent activation of the human sGC α 1/ β 1 is a promising approach to finding novel drugs or gene transfer techniques for cardiovascular therapy that neither result in tolerance in the patient nor form cytotoxic peroxynitrite. To find such pharmaceuticals, a mass screening for suitable active substances is necessary. Such a pharmacological screening for specific activators or inhibitors via animal testing is too expensive and is not reasonable due to species differences, possible side effects, and effects on other isoforms. Cell culture systems have the disadvantage that only with substantial additional effort can it be determined at which points of the signaling cascade substances are effective. In addition, cell culture is expensive and touchy. The purification of a protein from animal tissues is labor-intensive and results in a low yield. More importantly, because of species differences the results of such a screening are not generally transferable. Here, in particular, the question arises of the significance of the isoforms hsGC α 1 and hsGC α 3 and whether hsGC β 3 in fact

corresponds to a human hsGC β 1. This is an important factor to consider in selecting the appropriate target protein for pharmacological screening or gene therapy. For optimal screening, the human homologue of the protein sGC α 1/ β 1 would have to be purified in large amounts and to be available at low cost. A purification of large amounts of human native sGC α 1/ β 1 protein from human tissues is not possible. Thus, other methods are needed for the drug screening protocol. In addition, no purified antibodies are available for the detection of human sGC α 1/ β 1 that have been shown to be suitable for diagnostic uses, e.g. in normal immunoblotting, ELISA, RIA, or EIA, among other techniques. An approach for the artificial expression of hsGC by gene transfer in humans that could be used therapeutically is also not available.

Therefore, the present invention is based on the technical problem of providing isolated and purified human sGC α 1/ β 1 as well as a process for its production and purification. In addition, a further technical problem of the present invention is to provide antibodies directed against human sGC α 1/ β 1. A further technical problem is the provision of expression vectors containing the cDNA of human sGC α 1/ β 1 based on adenoviruses. Finally, the technical problem of the present invention is the provision of human sGC α 1/ β 1 in purified form and in manageable amounts for drug screening assays aimed at detecting modulators, inhibitors, and activators of human sGC α 1/ β 1.

The solution of the technical problem

The solution to the above technical problem is provided by the subject-matter of the claims and the following description of the invention.

An object of the invention is a human soluble guanylyl cyclase α 1/ β 1 (hsGC α 1/ β 1) in an isolated form purified to apparent homogeneity.

Another object of the invention is a process for the production of subunit α 1 and/or β 1 of human soluble guanylyl cyclase comprising the expression of expression vectors containing the DNA sequence for hsGC α 1 and/or hsGC β 1 in prokaryotic and eukaryotic host cells, and obtaining of the subunit or subunits.

In a preferred embodiment of the process of the present invention for the production of subunit $\alpha 1$ and/or $\beta 1$ of human soluble guanylyl cyclase, the step of obtaining the subunit or subunits comprises the lysis of cells, affinity chromatography of the cell lysate, and finally, the elution of the subunit or subunits.

In a further preferred embodiment of the process of the present invention for the production of subunit $\alpha 1$ and/or $\beta 1$ of human soluble guanylyl cyclase, the expression vector contains additionally at least one DNA sequence of a domain for specific affinity chromatography (affinity tag) with attached protease cleavage site.

Another object of the present invention is a process for the production of human soluble guanylyl cyclase $\alpha 1/\beta 1$ (hsGC $\alpha 1/\beta 1$) comprising the separate expression of an expression vector containing the DNA sequence for hsGC $\alpha 1$ or hsGC $\beta 1$ in prokaryotic or eukaryotic host cells, obtaining the subunits, and combining the subunits hsGC $\alpha 1$ and hsGC $\beta 1$ into the dimeric guanylyl cyclase $\alpha 1/\beta 1$ (hsGC $\alpha 1/\beta 1$).

A further preferred embodiment of the process of the present invention for the production of human soluble guanylyl cyclase $\alpha 1/\beta 1$ (hsGC $\alpha 1/\beta 1$) also comprises the coexpression of the DNA sequences for hsGC $\alpha 1$ and hsGC $\beta 1$ in prokaryotic or eukaryotic host cells, the lysis of the cells containing hsGC $\alpha 1$ and hsGC $\beta 1$, affinity chromatography, and the subsequent elution of hsGC $\alpha 1/\beta 1$.

Another object of the present invention is the use of a nucleic acid sequence that codes for subunits hsGC $\alpha 1$ and/or hsGC $\beta 1$ of human soluble guanylyl cyclase $\alpha 1/\beta 1$ for somatic gene therapy, particularly for the prevention and therapy of atherosclerosis and its complications, restenosis, ischemia (infarction), peripheral occlusive diseases, and arterial hypertension, as well as, in patients with risk factors for atherosclerosis.

In a particularly preferred embodiment, adenoviral vectors containing hsGC $\alpha 1$ or hsGC $\beta 1$ cDNA are used in somatic gene therapy. Other vector systems, however, can also be applied for gene therapeutic, medicinal use.

The methods of gene transfer described in the present invention can also be used for gene transfer of human soluble guanylyl cyclase $\alpha 2$ (GeneBank: x63282) and of the human homologue of soluble guanylyl cyclase $\beta 2$ (from rat; GeneBank: m57507) as well as for other human soluble guanylyl cyclases.

Another object of the present invention relates to antibodies to human soluble guanylyl cyclase $\alpha 1/\beta 1$ (hsGC $\alpha 1/\beta 1$), obtainable by immunization of a mammal with hsGC $\alpha 1/\beta 1$, either of subunits $\alpha 1$ or $\beta 1$, or immunogenic peptide fragments thereof, as well as isolation of the antibodies.

The Figures show:

Figure 1 shows various possibilities of modulation of soluble guanylyl cyclase (CsGC- $\alpha 1$ /sGC- $\beta 1$). Normal activation is mediated by NO synthase (NOS) and NO. NO, however, reacts with oxygen radicals to form peroxynitrite (ONOO⁻), which is cytotoxic and only poorly activates sGC. NO can also be released by NO donors such as nitroglycerin or sodium nitroprusside. sGC can be directly activated by modulators of sGC (e.g. YC-1); alternatively, the activation by NO of sGC is potentiated by these modulators. Additionally, sGC can be overexpressed by use of gene transfer (e.g. using adenoviral vectors) or a pathologically low expression level of sGC can be compensated. Adenoviral (or other) vectors with mutated sGC that has a higher basal activity can also be used. Thus, an elevated cGMP level could be achieved permanently independent of NOS, NO, NO donors, or sGC modulators.

Figure 2 shows a schematic comparison of the bovine and rat sGC $\alpha 1$ subunits together with the published sequence of the human cDNA clone termed 'sGC $\alpha 3$ ' (Giulli et al., 1992). The bars represent the protein. 'N' represents the N-terminus and 'C' the C-terminus. The functional segments 'regulatory domain', 'sGC homology domain', and 'cyclase domain' of these proteins are marked with different patterns. The regions 'S1' and 'S2' for which no homologous regions are found in the bovine and rat sGC $\alpha 1$ proteins are marked in black.

Figure 3 shows a schematic illustration of the human sGC α 3 clone with the sequence errors published (Giuli et al., 1992). The cDNA is shown above: the bar represents the coding region of the cDNA, the lines left and right of the bar represent the 5' and 3' untranslated regions, respectively. 'S1' and 'S2' represent the regions that have no
 5 homology to the bovine and rat isoforms of sGC α 1 (see Fig. 2). The positions of the sequence errors are marked below: line a shows the nucleotide insertions, line b the deletion, and line c the exchanges. A base-pair (bp) scale is shown below. The sequence errors for each of the 3 lines a, b, and c are listed at the bottom: the letter specifies the type of the base concerned, and the number its position in the cDNA.

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Figure 4 shows the verification of the expression of human sGC α 1 (A) and sGC β 1 (B) in human tissues by means of PCR using cDNA libraries. Shown is a photo of an ethidium bromide-stained agarose gel under UV light with separated PCR products. The arrow on the left points to the specific product. The PCR primers are visible at
 15 the bottom of the photo. The tissues from which the cDNA libraries were produced are indicated at the top. No cDNA was added in the negative control, and in the positive control, plasmid containing the cDNA of hsGC α 1 was added.

Figures 5 and 6 show the baculovirus transfer vectors pVL1393 and pAcG2T, respectively (both without the hsGC cDNA), which were used for the construction of
 20 recombinant baculoviruses for the expression of human sGC α 1/ β 1 in Sf9 cells. The circular plasmid with the restriction sites (short names and position in base pairs), the gene for ampicillin resistance (Amp^R), the 'origin of replication' (ColE ori), the polyhedrin promotor, the glutathione-S-transferase sequence (only in Fig. 6), and the
 25 multiple cloning site (MCS) are shown at the top of the figures. Figure 5 shows the multiple cloning site with its unique restriction sites at the bottom. Figure 6 shows the multiple cloning site with the unique restriction sites as well as a thrombin cleavage site normal at the bottom.

Figure 7 shows the construction of the plasmids hsGC β 1-pVL1393 (without GST-tag) with the hsGC β 1 cDNA, which was used to obtain genetically modified, hsGC β 1-expressing baculovirus by homologous recombination. The procedure for the plasmid pAcG2T-hsGC β 1 (with GST-tag = glutathione-S-transferase cDNA from *Schistosoma japonicum*) is identical. A fragment was produced that carries an additional BamHI-

site at its 5' end by means of PCR with the primers A (bases 89-116 of the hsGC β 1 cDNA + BamHI site at its 5' end) and B (bases 692-711 of the hsGC β 1 cDNA [noncoding strand] with natural KpnI site). Due to the additional restriction sites, fragment 1 (PCR fragment with new BamHI site) and fragment 2 (hsGC β 1 cDNA from KpnI site to EcoRI site) could be inserted together into the BamHI and EcoRI sites of the plasmid pVL1393. Thus, the complete hsGC β 1 cDNA is under the control of the polyhedrin promotor (PHP).

Figure 8 shows the construction of the plasmid hsGC α 1-pVL1393 (without GST-tag) with the hsGC α 1 cDNA which was used to obtain genetically modified, hsGC α 1-expressing baculovirus by homologous recombination. The procedure for the plasmid pAcG2T-hsGC α 1 (with GST-tag = glutathione-S-transferase cDNA from *Schistosoma japonicum*) is identical. A fragment was produced that carries an additional BamHI site at its 5' end and a natural BsaAI site within the sequence by means of PCR with the primers C (bases 524-541 of the hsGC α 1 cDNA + BamHI site at its 5' end) and D (bases 1232-1249 of the hsGC α 1 cDNA [noncoding strand]). Due to the added restriction site, fragment 3 cut with BsaAI (PCR fragment with new BamHI site to the BsaAI site) and fragment 4 (hsGC α 1 cDNA from BsaAI site to EcoRI site) could be inserted together into the BamHI and EcoRI sites of the plasmid pVL1393. Thus, the complete hsGC α 1 cDNA is under the control of the polyhedrin promotor (PHP).

Figure 9 shows the verification of the expression of hsGC α 1/ β 1 in Sf9 cells, which were infected with the genetically modified viruses described above (with hsGC α 1 or hsGC β 1 cDNA; both without GST-tag = glutathione-S-transferase cDNA from *Schistosoma japonicum*). On the left (A) a Coomassie-stained 10% SDS-polyacrylamide gel is shown on which were loaded the cell homogenates that had been separated into pellet (P) and supernatant (S) by centrifugation (20,000 x g). 'Co' designates the control with noninfected Sf9 cells. ' α 1' designates Sf9 cells that were infected with viruses containing the hsGC α 1 cDNA, and ' β 1' designates Sf9 cells that were infected with viruses containing the hsGC β 1 cDNA. The position of hsGC α 1 and hsGC β 1 in the gel are marked (α 1 or β 1). On the right an immunoblot is shown with supernatant (S) and pellet (P) of the cell homogenate from Sf9 cells that were either noninfected (Co) or coinfecting with hsGC α 1 and hsGC β 1 baculoviruses

($\alpha 1 + \beta 1$). The peptide antibodies to hsGC $\beta 1$ described above (anti-hsGC $\beta 1$) were used in immunoblotting first (Fig. 9B. lines 1-4). Afterwards, the blot was redeveloped with the peptide antibodies to hsGC $\alpha 1$ (anti-hsGC $\alpha 1$; Fig. 9B, lanes 5-8), which additionally revealed the hsGC $\beta 1$ bands.

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Figure 10 shows the activity of guanylyl cyclase (formation of cGMP from GTP) in intact Sf9 cells that were coinfectd with the genetically modified baculoviruses containing hsGC $\alpha 1$ and hsGC $\beta 1$ cDNA described here (both without GST-tag). Shown is the content of cGMP in pmol per 10^6 cells with different treatments, indicated at the bottom of the figure. Sample 1 is untreated in both panels A and B. 1 mM IBMX (3-isobutyl-1-methylxanthine) was added to each of the other samples (top line: black crossbar). In the middle line, the concentration of SNP added to the samples is indicated in μ M. The bottom line shows the concentration of added YC-1 (A, left) or ODQ (B, right) in μ M.

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15 YC-1 = 3-(5'-hydroxymethyl-2'-furyl)-1-benzylindazol;
 ODQ = 1H-[1,2,4]oxadiazolo[4,3,-a]chinoxalin-1-on;
 SNP = sodium nitroprusside;
 cGMP = cyclic 3', 5'-guanosine monophosphate

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Figure 11 shows the activity of guanylyl cyclase (formation of cGMP from GTP) in homogenates of Sf9 cells that were coinfectd with the genetically modified baculoviruses containing hsGC $\alpha 1$ and hsGC $\beta 1$ cDNA described here (both without GST-tag). As shown at the top, (A), the soluble cell fraction (supernatant after centrifugation at 20,000 x g) was used, and at the bottom, (B), the respective pellet. In each case, the amount of cGMP formed in pmol per mg protein per minute is shown for homogenates of cells that were harvested at different time points (indicated in hours) after infection with the baculoviruses. cGMP formation was measured with (black boxes) and without (white boxes) the addition of 100 μ M SNP.

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Figure 12 shows the activity of guanylyl cyclase (formation of cGMP from GTP) in Sf9 cells that were coinfectd with the genetically modified baculoviruses containing hsGC $\beta 1$ cDNA (without GST-tag) and hsGC $\alpha 1$ cDNA (with GST-tag = glutathione-S-transferase cDNA from *Schistosoma japonicum*). The formation of cGMP in pmol per mg protein per minute during the purification procedure (affinity chromatography on

glutathione sepharose 4B) is shown. In each case, activity was measured in the lysate (after removing the insoluble part by centrifugation at 20,000 x g), in the supernatant after binding of hsGC to glutathione sepharose 4B (flow-through), in both supernatants of the washes of hsGC bound to glutathione sepharose 4B (1. and 2. wash), as well as in the supernatant after elution of hsGC with reduced glutathione (1. and 2. elution). The formation of cGMP was determined without (black boxes, 'basal') and with (grey boxes, '+ 100 μ M SNP') addition of 100 μ M SNP.

Figure 13 shows the natural, endogenous expression of hsGC α 1 and hsGC β 1 in different human tissues in an immunoblot.

The peptide antibodies to hsGC α 1 described above (anti-hsGC α 1) were used on the left (A), and the peptide antibodies to hsGC β 1 (anti-hsGC β 1) were used on the right (B). On the right, the peptide to which the antibodies were raised was added as negative control (peptide: +), while no peptide was added on the left (peptide: -). The SDS extracts of rhsGC α 1- (in panel A of the figure) or rhsGC β 1-overexpressing Sf9 cells (in panel B of the figure) (Sf9), of human cerebral cortex (cortex), of human cerebellum (cerebellum), and of human lung (lung) were loaded on an 8% polyacrylamide gel. The specific bands of hsGC α 1 (α 1) and hsGC β 1 (β 1) are indicated by an arrow.

Figure 14 shows in immunoblots the purification of hsGC α 1 (as a hsGC α 1/hsGC β 1 dimer) from Sf9 cells that were coinfecting with the genetically modified baculoviruses containing hsGC α 1 cDNA with GST-tag (= glutathione-S-transferase cDNA from *Schistosoma japonicum*) and hsGC β 1 cDNA (without GST-tag). Cell lysate was incubated with glutathione sepharose 4B, and after binding, the supernatant was loaded (supernatant after binding). The sepharose was washed twice and the respective supernatants of this wash were loaded (1. and 2. wash). Subsequently, elution was performed by cleavage of the hsGC α 1 protein from the GST-tag with thrombin and an aliquot was loaded ('elution with thrombin'). Then, SDS stop buffer was added to the glutathione sepharose 4B and an aliquot was loaded (GSH sepharose after elution). In addition, glutathione sepharose 4B with bound hsGC α 1 without prior thrombin elution to which SDS buffer had been added was loaded (GSH sepharose before elution). The immunoblot was developed with the affinity-purified peptide antibodies to the C-terminus of hsGC α 1 described here. The arrows on the

right point to the specific bands of hsGC α 1 with the GST-tag (GST-hsGC α 1) and hsGC α 1 without the GST-tag (hsGC α 1).

Figure 15 shows the purification of hsGC α 1/ β 1 from Sf9 cells that were coinfectd with the genetically modified baculoviruses containing hsGC α 1 cDNA with GST-tag (= glutathione-S-transferase cDNA from *Schistosoma japonicum*) and hsGC β 1 cDNA without GST-tag in a Coomassie Brilliant Blue R250-stained SDS-polyacrylamide gel. Cell lysate of these infected Sf9 cells (lysate) was incubated with glutathione sepharose 4B, and the supernatant loaded after the binding (supernatant after binding). The glutathione sepharose 4B was washed twice and the respective supernatants of this wash buffer were loaded (1. and 2. wash). In one sample, the bound GST-hsGC α 1/ β 1 was eluted by incubation with reduced glutathione and loaded (elution with glutathione). In the other samples, the glutathione sepharose was washed with the buffer for the thrombin cleavage—without thrombin—and the supernatant of this buffer was loaded (3. wash). Then, the hsGC α 1/ β 1 dimer was eluted by incubation with different amounts of thrombin and the eluates were loaded (elution with 0.25-1 U/ml thrombin) [U = unit]. The same relative amount of each of the samples was used. The bands visible with the different elution methods are indicated on the right: GST-hsGC α 1 = hsGC α 1 with GST-tag; hsGC α 1 = hsGC α 1 without GST-tag; hsGC β 1 = hsGC β 1 without GST-tag. On the left are the molecular weight standards loaded on the gel, the size of which (in kDa) is indicated on the far left side.

Figure 16 shows the construction of the recombinant adenoviral hsGC vectors. The cDNAs of hsGC α 1 and hsGC β 1 (grey bars) were inserted in the adenoviral transfer plasmid pZS2, which has a deletion in the adenovirus E1 region (Δ E1) and a unique XbaI site in this plasmid. This resulted in the plasmids hsGC α 1-pZS2 and hsGC β 1-pZS2, respectively. hsGC α 1-pZS2 and hsGC β 1-pZS2 cut with the restriction enzyme XbaI (middle bar, indicated as 'sGCpZS2') were ligated into the XbaI site of the long arm (upper bar, 'RR5') of Ad5. This resulted in the adenovectors Ad5CMVhsGC α 1 and Ad5CMVhsGC β 1, respectively (lower bar, 'Ad 5 CMV sGC') in which the sGC cDNAs lie under the control of the CMV promotor and CMV enhancer (CMV = cytomegalovirus).

Figure 17 shows the stimulation of sGC activity by 100 μ M SNP (= sodium nitroprusside) in EA.hy926 cells that were coinfectd with both hsGC adenoviruses Ad5CMVhsGC α 1 and Ad5CMVhsGC β 1 (samples A-C) and in noninfected EA.hy926 cells (sample D). The amount of pmol cGMP formed per mg protein per minute is plotted on the Y axis. The dark bars represent basal cGMP formation without SNP stimulation, and the light bars represent cGMP formation with SNP stimulation.

Figure 18 shows the DNA sequence of human soluble guanylyl cyclase α 1 (hsGC α 1); SEQ ID NO: 1.

Figure 19 shows the amino acid sequence of human soluble guanylyl cyclase α 1 (hsGC α 1); SEQ ID NO: 2.

Figure 20 shows the DNA sequence of human soluble guanylyl cyclase β 1 (hsGC β 1); SEQ ID NO: 3.

Figure 21 shows the amino acid sequence of human soluble guanylyl cyclase β 1 (hsGC β 1); SEQ ID NO: 4.

Figure 22 shows the amino acid sequence of the peptide that was used for the production of antibodies to human soluble guanylyl cyclase α 1 (hsGC α 1) (corresponds to amino acids 634-647 of hsGC α 1); SEQ ID NO: 5.

Figure 23 shows the amino acid sequence of the peptide that was used for the production of antibodies to human soluble guanylyl cyclase β 1 (hsGC β 1) (corresponds to amino acids 593-614 of hsGC β 1); SEQ ID NO: 6.

Figure 24 shows the DNA sequence of the PCR primer pair for human soluble guanylyl cyclase α 1 (hsGC α 1). Upper primer (corresponds to nucleotides 524-541 of the hsGC α 1 cDNA sequence with added BamHI restriction site); SEQ ID NO: 7. Lower primer (corresponds to nucleotides 1249-1232 of the hsGC α 1 cDNA sequence [noncoding strand]); SEQ ID NO: 8.

Figure 25 shows the DNA sequence of the PCR primer pair for human soluble guanylyl cyclase $\beta 1$ (hsGC $\beta 1$). Upper primer (corresponds to nucleotides 89-106 of the hsGC $\beta 1$ cDNA sequence with added BamHI restriction site); SEQ ID NO: 9.

- 5 Lower primer (corresponds to nucleotides 692-711 of the hsGC $\beta 1$ cDNA sequence [noncoding strand]); SEQ ID NO: 10.

Advantages of the present invention and solution of the above-mentioned technical problems according to the present invention

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1.) The cDNA clones described in the literature as sGC $\alpha 3$ and sGC $\beta 3$ were identified as human homologues of bovine and rat sGC $\alpha 1$ and sGC $\beta 1$, and in the following are termed human sGC $\alpha 1$ (hsGC $\alpha 1$) and human sGC $\beta 1$ (hsGC $\beta 1$). According to present knowledge, this sGC $\alpha 1/\beta 1$ isoform is pharmacologically more important due to its function in the cardiovascular system. Because the original clone of hsGC $\alpha 3$ was examined, it could be shown that hsGC $\alpha 1$ and hsGC $\alpha 3$ do not exist in parallel but rather only the form hsGC $\alpha 1$. Thus, an obvious target protein for pharmacological mass screening and gene therapy has been identified.

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2.) With the methods described by the present invention, a functional, active expression of human sGC has been obtained for the first time. Thus, the respective protein can be produced by genetic methods for the first time.

25 3.) By use of the peptide antibodies to sGC of the present invention, it is possible to determine the expression of sGC in human tissues as well as diagnose dysfunctional conditions (if expression of sGC is too high, too low, or absent). In addition, the present invention provides the technical prerequisites needed to further elucidate the control of transcription and translation of hsGC. The peptide antibodies of the invention have the advantage that they are monospecific, directed at the human sequence, and that their suitability for immunoblots with human tissues has been demonstrated. Other peptide antibodies exhibit these features only partially: Harteneck et al. and Guthmann et al. used a peptide sequence (VYKVETVGDKYMTVSGLP) that is relatively highly conserved in

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guanylyl cyclases. Thus, cross-reaction with particulate guanylyl cyclases (e.g. GC-C) would be expected. Furthermore, Guthmann et al. used a peptide sequence (YGPEVWEDIKKEA) identical to hsGC β 1 and a peptide sequence identical to hsGC α 1 except for two amino acid exchanges (KKDVEEANANFLGKASGID), but the function of these antibodies in immunoblotting has only been shown for enriched hsGC from human platelets. In addition, these antibodies to hsGC α 1 recognized a second, unspecific product. Humbert et al. and Koesling et al. used a peptide sequence (SRKNTGTEETE QDEN) from bovine sGC β 1 that is in part (amino acids 1-10) identical to the peptide used here (amino acids 13-22) for hsGC β 1, although the C-terminus (amino acids 11-15) differed markedly from that of the human sequence. The antiserum to this peptide, however, has not been tested on human protein and has only been used for immunoprecipitation of bovine sGC.

In addition to the peptides shown in Figures 22 and 23 and their immunogenic fragments for the production of the antibodies to hsGC α 1 or hsGC β 1 in rabbits of the present invention, the production of monoclonal or polyclonal antibodies to the whole hsGC α 1/ β 1 protein or its cleavage products is possible. Various animal species (preferably mouse, rat, or rabbit) can be used for the production of these antibodies.

4.) By use of the eukaryotic baculovirus/Sf9 expression system of the present invention, human soluble guanylyl cyclase α 1/ β 1 can be produced in high amounts. The attachment of a nucleotide sequence that codes for a polypeptide suitable for affinity chromatography (affinity tag, e.g. glutathione-S-transferase = GST-tag) with an attached protease cleavage site at the N-terminus of the cDNA of the α 1 subunit allows the rapid and simple purification of the coexpressed dimeric protein by means of a single affinity chromatographic step. The attached affinity tag is subsequently removed by digestion with a protease. Thus, a protein identical in primary structure to the native protein is obtained. This revolutionary, fast, and clean purification yielding high amounts of very pure, functional human sGC results in new possibilities for a mass screening for specific activators and inhibitors as well as for the pharmacological characterization of potential drugs.

Purification of hsGC α 1/ β 1 can also be accomplished by ion exchange chromatography, gel filtration, immunoaffinity chromatography, and other chromatographic procedures, e.g. on ATP-, GTP-, cGMP- or Blue-sepharose, and other similar chromatographic media.

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5.) The process of the present invention can also be used for other isoforms of the human, rat, and bovine enzyme in an identical manner. In the process provided in the present invention, various affinity tags (e.g. histidine oligomer) and different expression systems (e.g. *E. coli*) can be used. Other parts of the hsGC α 1 and hsGC β 1 sequences can also be used for the production of antibodies to peptides or the whole protein.

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6.) The availability of high amounts of an isolated human protein in high purity and in high quality is essential for modern pharmaceutical development. Until now, this requirement was not fulfilled in the search for alternatives to the classical NO donors. Mass screening for specific activators or inhibitors in animal testing is too expensive and would not make sense due to species differences, possible side effects, and effects on other isoforms. Cell culture systems have the disadvantage of requiring substantial additional effort to determine precisely at which point within the signaling cascades substances act. In addition, cell culture is expensive and touchy. Compared with the expression of a protein using recombinant DNA technology, the purification of a protein from animal tissues is more labor intensive and results in lower yields. In particular, the results of a pharmacological screening can only be generalized to a limited extent because of differences between species. In contrast, processes of the present invention provide for recombinant, inexpensive hsGC α 1/ β 1 in high amounts, and unequivocal predictions concerning modulation, activation, or inhibition of this particular human enzyme can be made from a screening procedure. Species differences and lack of applicability to human are excluded a priori by the use of a human enzyme.

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In addition, hsGC is available thereby in high amounts and adequate purity for crystallization and clarification of its structure. Thus, an important prerequisite for rational drug design by means of molecular modeling is fulfilled.

7.) In addition to the use of isolated hsGC α 1/ β 1, intact Sf9 cells that express hsGC α 1/ β 1 by being infected with the recombinant baculoviruses described here can be used in in vitro experiments.

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8.) Transient (e.g. with adenoviral vectors) or stable overexpression can be achieved by means of gene transfer. Thus, the cGMP level can be elevated even at low NO concentration or in the case of poor activation of sGC due to peroxynitrite formation. This approach also has conceptual advantages compared to the gene transfer of NOS (NO synthase) because the formation of cytotoxic peroxynitrite, which is less effective in activating sGC, is circumvented with the approach of the present invention. Furthermore, a permanent increase in the cGMP level could be achieved (e.g. for therapeutic purposes) independently of NOS, NO, NO donors, or sGC modulators by gene transfer of mutated sGC with elevated basal activity.

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9.) In addition to the purification procedure for hsGC α 1/ β 1 described here, purification from Sf9 cells is also possible after infection with the described baculoviruses containing hsGC α 1 or hsGC β 1 cDNA if coinfection is performed with hsGC α 1 baculoviruses without a GST-tag (GST-tag = appended glutathione-S-transferase sequence from *Schistosoma japonicum*) and hsGC β 1 baculoviruses having a GST-tag as well as with hsGC α 1 baculoviruses having a GST-tag and hsGC β 1 baculoviruses having a GST-tag.

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Furthermore, use of a column containing glutathione sepharose 4B is possible in addition to the use of glutathione sepharose 4B alone in a batch-wise step.

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Purification can also be performed by digesting the dimeric fusion protein GST-hsGC α 1/ β 1 with thrombin *after* elution of the fusion protein from the glutathione sepharose 4B with reduced glutathione. After dialysis (to remove the reduced glutathione), the GST-tag which has been cleaved from the protein can be removed from the mixture by additional affinity chromatography on glutathione sepharose 4B.

10.) The process of the present invention can also be used for human soluble guanylyl cyclase $\alpha 2$ (GeneBank: x63282) and any potentially existing human homologue of the soluble guanylyl cyclase $\beta 2$ (from rat; GeneBank: m57507) as well as for other human soluble guanylyl cyclases (in all technical variations described in the present invention).

The examples illustrate the present invention.

Example 1

The corrected sequence of hsGC $\alpha 1$ and hsGC $\beta 1$

The original clone of the human isoform sGC $\alpha 3$ and sGC $\beta 3$ (Giulii et al., 1992) was sequenced again. While the sequence of the sGC $\beta 3$ clone was confirmed (see SEQ ID NO: 3 and Figure 20), the sequencing of sGC $\alpha 3$ showed that the original publication (Giulii et al., 1992) contained 19 sequencing errors, which are summarized in Figure 3. The corresponding corrected $\alpha 3$ -cDNA sequence is shown in SEQ ID NO: 1 and Figure 18. The deduced amino acid sequence is shown in SEQ ID NO: 2 and Figure 19. Furthermore, the corrected sequence (see SEQ ID NO: 1 and Figure 18) is identical with the human sGC $\alpha 1$ sequence published in the GeneBank (accession No. U58855), whereby the 5' untranslated region of the sequence provided here is 506 base pairs longer. Therefore, 'sGC $\alpha 3$ ' is now classified as human sGC $\alpha 1$ (hsGC $\alpha 1$). Thus, it was shown that two different hsGC α subunits $\alpha 1$ and $\alpha 3$ do not both exist in humans, which could be important for cardiovascular research but rather—in analogy to the situation in bovine and rat tissues—only hsGC $\alpha 1$.

Table 1 Revised terminology of soluble guanylyl cyclase cDNAs and proteins and their detection in human tissue.

human sGC subunits		
	α	β
isoform 1	cDNA and protein detectable	cDNA and protein detectable

active, when coexpressed

isoform 2

cDNA detectable

active, when coexpressed with bovine $\beta 1$

The expression of sGC $\alpha 1$ and sGC $\beta 1$ mRNA in human tissues was demonstrated by means of PCR (Figure 4). The amplification of a hsGC $\beta 1$ fragment with a PCR primer pair (5'-AAAAGGATCCATGTACGGATTTGTGAAT-3' = nucleotides 89-106 of the
5 hsGC $\beta 1$ cDNA sequence with added restriction site; 5'-ATGCGTGATTCCTGGGTACC-3' = 692-711 of the hsGC $\beta 1$ cDNA sequence) with an annealing temperature of 54°C resulted in one specific band each in cDNA libraries from brain, heart, kidney, lung, pancreas, and skeletal muscle. The identity of the amplified fragment was confirmed by sequencing. The amplification of a
10 hsGC $\alpha 1$ fragment with a PCR primer pair (5'-AAAAGGATCCATGTTCTGCACGAAGCTC-3' = nucleotide 524-541 of the hsGC $\alpha 1$ cDNA sequence with added restriction site; 5'-ATTATGGAAGCAGGGAGG-3' = 1249-1232 of the hsGC $\alpha 1$ cDNA sequence) with an annealing temperature of 54°C resulted in one specific band each in cDNA libraries from heart (Figure 4A) and lung
15 (not shown). In each case, the sequencing of the fragments resulted in the corrected hsGC $\alpha 1$ sequence; and the 'hsGC $\alpha 3$ ' sequence published by Giulli et al. was not found. Thus, it was shown that in humans, only one hsGC $\alpha 1/\beta 1$ exists, and that the potential hsGC $\alpha 3/\beta 3$ is a result of sequencing errors. This results in a clear picture for cardiovascular research concerning which sGC isoform should be the target
20 protein for pharmacological screening.

Example 2

25 *Construction of recombinant baculoviruses for the expression of human sGC α and sGC β in insect cells*

In order to verify that hsGC $\alpha 1$ and hsGC $\beta 1$ can form a functional, heterodimeric sGC protein, both cDNAs were inserted into baculoviruses. Using these baculoviruses, the

recombinant protein was expressed under the control of the strong polyhedrin promoter in insect cells (Sf9 cells). For the production of the recombinant baculoviruses, the baculovirus transfer vector pVL1393 (Pharmingen, San Diego, California, USA; Figure 5) and the baculovirus transfer vector pAcG2T (with glutathione-S-transferase sequence from *Schistosoma japonicum* and thrombin cleavage site; Pharmingen; Figure 6) were used in which the foreign genes hsGC α 1 and hsGC β 1 were cloned, respectively. The cotransfection of such a recombinant pVL1393 or pAcG2T plasmid with BaculoGold baculovirus DNA (Pharmingen) allowed the direct isolation of the genetically modified baculoviruses with hsGC α 1 or hsGC β 1 cDNA formed by homologous recombination from the cell culture media.

The construction of pVL1393-hsGC β 1 is schematically shown in Figure 7 (identical procedure for pAcG2T-hsGC β 1). The coding region of the hsGC β 1 cDNA with the 3' untranslated region but without the 5' untranslated region was cloned into pVL1393. For this, a BamHI site was introduced by means of PCR with primers A and B immediately upstream of the codon coding for the start methionine. The thus amplified fragment 1 was digested with BamHI/KpnI; fragment 2 was isolated from the sGC β 1 cDNA clone with KpnI/EcoRI. Fragments 1 and 2 as well as the vector opened with BamHI and EcoRI were ligated (see Figure 7).

The construction of pVL1393-hsGC α 1 is shown schematically in Figure 8 (identical procedure for pAcG2T-hsGC α 1). The coding region of the hsGC α 1 cDNA with the 3' untranslated region but without the 5' untranslated region was cloned into pVL1393. For this, a BamHI site was introduced by means of PCR with primers C and D immediately upstream of the codon coding for the start methionine. The thus amplified fragment 3 was digested with BamHI/BsaAI; fragment 4 was isolated from the sGC α 1 cDNA clone with BsaAI/EcoRI. Fragments 3 and 4 as well as the vector opened with BamHI and EcoRI were ligated (see Figure 8).

For the production of the recombinant hsGC α 1 and hsGC β 1 baculoviruses, the baculovirus transfer vectors (pVL1393-hsGC α 1, pAcG2T-hsGC α 1, pVL1393-hsGC β 1, pAcG2T-hsGC β 1) were each cotransfected with baculovirus DNA (BaculoGold; Pharmingen, San Diego, California, USA) in monolayers of Sf9 cell cultures.

For this, the cells were cultivated at 27°C in IPL-41 media (Gibco) supplemented with 10% (vol/vol) fetal calf serum (Biochrom), 4% (vol/vol) tryptose-phosphate-broth (Gibco), 1% (vol/vol) Pluronic F68 (Gibco), 0,5% amphotericin B (Gibco), 80 µg/ml gentamycin sulfate (Gibco), and 0.5 mM δ -aminolevulinic acid (Merck). Recombinant
 5 hsGC α 1 and hsGC β 1 baculovirus clones were obtained from the culture media by means of plaque purification. For the production of virus stock solutions with high titer, Sf9 shaking cultures (0.5×10^6 cells/ml) were infected with a M.O.I. (multiplicity of infection) of 0.1 pfu/cell (pfu = plaque forming units) and harvested 6 days after infection.

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Example 3

Production of recombinant hsGC α 1 and hsGC β 1 in Sf9 cells

Ten of each recombinant hsGC α 1 and hsGC β 1 baculovirus clones were tested for
 15 expression of recombinant protein in Sf9 cells. For this, Sf9 monolayer cell cultures were infected with plaque-purified, recombinant hsGC α 1 or hsGC β 1 baculoviruses, incubated at 27°C for 5 days, harvested with a cell scraper, resuspended in 0.5 ml lysis buffer (25 mM TEA, pH 7.8, 50 mM NaCl, 1 mM EDTA, 5 mM DTT, 1 µM leupeptin, 0.5 mg/l trypsin inhibitor), and homogenized by sonication ('Sonifier 250',
 20 'standard'-tip, Branson; 15 times, 'duty cycle': 15%, intensity: 1). After centrifugation of the homogenates at 20,000 x g, the supernatant and pellet were analyzed by SDS-PAGE. Three of the hsGC β 1 baculovirus clones and two of the hsGC α 1 baculovirus clones yielded recombinant protein in amounts that were visible in the insoluble fraction by staining with Coomassie Brilliant Blue R250. Recombinant human sGC α
 25 and sGC β (rhsGC α and rhsGC β) migrated with an apparent molecular weight of 79.5 (hsGC α 1) and 68.5 kDa (hsGC β 1), which is very close to the predicted molecular weights deduced from the amino acid sequence (77.5 and 70.5 kDa, respectively) (representative clones in Figure 9). The baculovirus clones that showed the highest expression of recombinant proteins in the immunoblot were used for the expression
 30 of functional heterodimeric hsGC [see examples 4-7].

Example 4

Recombinant human sGC in intact insect cells is active and stimulated by NO

For the production of functional heterodimeric human sGC, rhsGC α 1 and rhsGC β 1 were coexpressed in Sf9 cells with recombinant baculoviruses. For this, Sf9 monolayer cultures [2.5×10^6 cells/dish, Ø 90 mm; supplements see example 2] were
 5 coinfectd with a M.O.I. (multiplicity of infection) of 2 pfu/cell of each recombinant baculovirus (hsGC α 1 and hsGC β 1; both without GST-tag) and cultivated for 48 hours at 27°C. The basal as well as the NO-stimulated activity of the sGC in the cells was determined by measurement of the cGMP content in the cells in the presence of the phosphodiesterase inhibitor IBMX (3-isobutyl-1-methylxanthine).

10 For the determination of the cGMP content, the culture media was replaced by Krebs-Ringer-buffer (KRB; 119 mM NaCl; 4.74 mM KCl; 2.54 mM CaCl₂; 1.19 mM MgSO₄; 1.19 mM KH₂PO₄; 25 mM NaHCO₃; 10 mM HEPES; pH 7.4; 0.1% BSA) which, in addition, contained 1 mM IBMX. The cells were incubated for one hour at
 15 27°C. Then the cells were washed with ice-cold KRP and harvested in 1 ml ice-cold ethanol (80 %) with a cell scraper. The cells were homogenized by sonication [see Example 3] and centrifuged at 20,000 x g for 20 minutes. The supernatant was dried in a speed-vac and the residue resuspended in 25 mM TEA, pH 7.8. The cGMP content was determined by means of RIA (Biotrend).

20 The coexpression of rhsGC α 1 and rhsGC β 1 resulted in the formation of functional sGC with basal activity in Sf9 cells (Figure 10): While noninfected Sf9 cells contained approximately 0.1 pmol cGMP/ 10^6 cells (not shown), approximately 20 pmol cGMP/ 10^6 cells was found in rhsGC-expressing cells (Figure 10). This basal activity
 25 of recombinant hsGC was increased by an NO donor, SNP (sodium nitroprusside). When the cells were incubated with 10, 100, or 1000 μ M SNP for 2 minutes prior to the harvest, the cGMP content increased in a concentration-dependent manner by up to 50 fold (Figure 10).

Example 5

Recombinant human sGC in insect cell extracts is active and stimulated by NO

The activity of the recombinant hsGC (after expression with the recombinant baculoviruses described above) was not only determined in intact Sf9 cells but also in Sf9 cell extracts. For the production of such extracts, Sf9 shaking cultures [2×10^6 cells/ml; supplements see Example 2] were coinfectd with a M.O.I. (multiplicity of infection) of 1 pfu/cell of each virus (hsGC α 1 and hsGC β 1; both without GST-tag) and incubated at 27°C. Samples were taken (4 ml) at 0, 24, 48, 72, 96, and 118 hours after infection, the cells sedimented, resuspended in 1 ml lysis buffer and homogenized by sonication [see Example 3]. The homogenates were centrifuged at 20,000 x g for 15 minutes and the insoluble pellet was resuspended again in lysis buffer. The samples were adjusted to 50% glycerin (vol/vol) and stored at -20°C. The protein concentration was determined spectrophotometrically with the standard method of Bradford (Bradford, 1976). The sGC activity was determined by the formation of [32 P]cGMP from [α - 32 P]GTP (Schultz and Böhme, 1984). The reactions contained 50 mM TEA (pH 7.4), 3 mM MgCl₂, 3 mM DTT, 1 mM IBMX, 1 mM cGMP, 5 mM creatine phosphate, 0.25 mg/ml creatine kinase, and 500 μ M GTP in a total volume of 100 μ l. After incubation at 37°C for 10 minutes, the reaction was started by simultaneous addition of cell extract and the sGC activators SNP, CO, or YC-1. The [32 P]cGMP formed was measured as described (Schultz and Böhme, 1984). Basal rhsGC activity (i.e. cGMP formation by rhsGC *without* activation of the enzyme by addition of NO or other activators), mainly found in the soluble Sf9 cell fraction (Figure 11A), reached its maximum 72 hours after infection of the cells and was increased up to 5 fold with 100 μ M SNP (Figure 11A). The pellet fraction did not contain measurable basal sGC activity at any time point; although in the presence of SNP a low degree of sGC activity was found (Figure 11B).

Example 6

Influence of YC-1 and ODQ on recombinant human sGC

YC-1 (3-(5'-hydroxymethyl-2'-furyl)-1-benzylindazol) and ODQ (1H-[1,2,4]oxadiazol[4,3,-a]chinoxalin-1-on) are substances that were described to specifically influence the activity of sGC. Thus, it was investigated if this also holds true for rhsGC.

After expression with the recombinant baculoviruses (without GST-tag) described above, rhsGC was activated in intact Sf9 cells by YC-1. The NO-potentiating effect was also found: the cGMP content of cells expressing rhsGC was increased 3.4 fold by incubation with 10 μ M YC-1 for 2 minutes (Figure 10A). 100 μ M YC-1 had the same effect (Figure 10A). When the cells were simultaneously treated with YC-1 and 100 μ M SNP, the cGMP levels doubled compared to the cGMP levels after stimulation with SNP alone (Figure 10A). Similar results were obtained with rhsGC in cell extracts.

ODQ is described as a selective inhibitor of NO-stimulated sGC that, however, does not inhibit the basal activity (Garthwaite et al., 1995). In Sf9 cells expressing rhsGC (after expression with the recombinant baculoviruses described above, without GST-tag), ODQ did not have an influence on the basal cGMP levels; the stimulation of rhsGC in intact cells with SNP, however, was inhibited by simultaneous incubation with ODQ (Figure 10B).

Example 7

Extraction of the purified human soluble guanylyl cyclase $\alpha 1/\beta 1$ (hsGC $\alpha 1/\beta 1$)

For the purification of recombinant human hsGC $\alpha 1/\beta 1$ from Sf9 cells, a recombinant baculovirus [see Example 2] was used in which a fusion protein composed of hsGC $\alpha 1$ and attached GST [so-called GST-tag, GST = glutathione-S-transferase from *Schistosoma japonicum*; see Example 2] is formed. Using this GST-tag, which binds to glutathione with high affinity, specific affinity chromatography on glutathione sepharose 4B (Pharmacia, Freiburg, Germany) can be performed. The Sf9 cells coinfecting with hsGC $\alpha 1$ baculoviruses (with GST-tag) and hsGC $\beta 1$ baculoviruses (without GST-tag) were lysed in 25 mM triethanolamine pH 7.8 / 1 mM EDTA / 5 mM DTT / 1 μ M leupeptin / 0.5 μ g/ml trypsin inhibitor / 0.2 mM PMSF (30 min hypotonic lysis at 4°C). After addition of NaCl (75 mM final concentration), the homogenate was centrifuged at 75,000 x g for 1 hour at 4°C. The supernatant was mixed with the GSH sepharose 4B for 1 hour at room temperature. The glutathione sepharose 4B was then pelleted by centrifugation at 500 x g for 5 minutes and the supernatant removed. A 10-fold volume of 50 mM Tris-HCl (pH 8.0) / 150 mM NaCl / 2.5 mM CaCl₂ / 0.1% 2-mercaptoethanol was added to the glutathione sepharose 4B and mixed for 1 minute. The mixture was centrifuged again at 500 x g for 5 minutes. The supernatant

was removed and the glutathione sepharose 4B was washed again in the same way. To elute from the sepharose, the hsGC α 1 protein (with the bound hsGC β 1) was cleaved by thrombin from the GST-tag, which remained bound to the glutathione sepharose 4B at the specific cleavage site. Digestion with thrombin was performed in
 5 50 mM Tris-HCl (pH 8.0) / 150 mM NaCl / 2.5 mM CaCl₂ / 0.1% 2-mercaptoethanol with 0.25 to 1 units thrombin/ml buffer for 1 or 3 hours at room temperature. The glutathione sepharose 4B (with the GST-tag) was pelleted again by centrifugation at 500 x g for 5 minutes and the supernatant containing the hsGC α 1/ β 1 was removed. Another elution method was performed by addition of 50 mM Tris-HCl (pH 8.0) / 5
 10 mM reduced glutathione and mixing for 30 minutes at room temperature. In this manner, hsGC α 1/ β 1 with the GST-tag bound was removed from the glutathione sepharose 4B. After centrifugation at 500 x g for 5 minutes, the supernatant containing dissolved GST-hsGC α 1/ β 1 was removed.

15 By the elution with thrombin, a two-fold selectivity is achieved in a single affinity chromatographic step:

- 1.) Only proteins that have an affinity to reduced glutathione are able to bind.
- 2.) Of these proteins, only the proteins that are cleaved by thrombin will elute (as cleavage products).

20 The thrombin can be separated from the sample with a p-aminobenzamidine column to which thrombin binds specifically.

Figure 12 shows the specific enrichment of sGC activity after elution from the GSH
 25 sepharose 4B with glutathione compared with the activity in the lysate of infected Sf9 cells.

Figure 14 shows the binding of GST-hsGC α 1 to glutathione sepharose 4B and the cleavage of hsGC α 1 from the GST-tag by thrombin in an immunoblot.

30 Figure 15 shows the purification of coexpressed hsGC α 1 with GST-tag and hsGC β 1 without GST-tag by affinity chromatography on glutathione sepharose 4B in a Coomassie Brilliant Blue R250-stained SDS-polyacrylamide gel. Upon elution with reduced glutathione, only two bands were visible, which correspond to GST-hsGC α 1

(larger product) and hsGC β 1 (smaller product) (detected in an immunoblot). After elution with thrombin (0.25, 0.5, or 1 unit/ml for three hours at room temperature), however, the lower band was identical (hsGC β 1; estimated molecular weight according to the migration in the gel of approximately 70 kDa), whereas the upper band was significantly smaller compared to elution with glutathione (hsGC α 1; estimated molecular weight of approximately 80 kDa) because the GST-tag was cleaved off by use of the thrombin elution. This approximately corresponds to the molecular weights of 77.5 kDa for hsGC α 1 and 70.5 kDa for hsGC β 1 deduced from the amino acid sequences. In contrast to the elution with reduced glutathione, an additional very small band of approximately 25 kDa was visible after the thrombin elution, which is likely thrombin itself. Thrombin can be removed from the eluate by means of an aminobenzamidine sepharose column to which thrombin binds specifically. Additional bands were not detectable in this experiment.

Example 8

Production of polyclonal rabbit antisera to hsGC α 1 and hsGC β 1

Antisera were obtained by immunization of rabbits with synthetic peptides corresponding to sequences from hsGC α 1 (Phe-Thr-Pro-Arg-Ser-Arg-Glu-Glu-Leu-Pro-Pro-Asn-Phe-Pro [Figure 22/SEQ ID NO: 5]; amino acids 634-647) and from hsGC β 1 (Lys-Gly-Lys-Lys-Glu-Pro-Met-Gln-Val-Trp-Phe-Leu-Ser-Arg-Lys-Asn-Thr-Gly-Thr-Glu-Glu-Thr [Figure 23/SEQ ID NO: 6] amino acids 593-614) that were coupled to KLH (keyhole limpet hemocyanin) via an additional C-terminal (α 1) or N-terminal (β 1) cysteine residue. The antisera were affinity-purified with the corresponding peptides coupled to epoxy-activated sepharose (Pharmacia, Freiburg, Germany) according to the manufacturer's instructions.

Example 9

Detection of hsGC α 1 and hsGC β 1 in different human tissues by immunoblotting

Human lung tissue was obtained from a tumor-free area of a lung resection, and human cortex and cerebellum were from a normal autopsy. All tissues were immediately frozen in liquid nitrogen and stored at -70°C . The frozen tissues were homogenized in a mortar and double-concentrated, hot SDS stop buffer (130 mM

Tris-HCl, pH 6.8 / 16% [v/v] glycerol / 4% [w/v] SDS / 0.025% [w/v] bromphenol blue / 6.5% [v/v] 2-mercaptoethanol) was added to the powder. This was incubated at 95°C for 10 minutes and then centrifuged at 20,000 x g for 20 minutes. The supernatant was used for immunoblotting (for antibodies used, see above).

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Expression of both subunits ($\alpha 1$ and $\beta 1$) was detected in all three tissues (Figure 13). In contrast, expression could not be detected in kidney, liver, and pancreas (data not shown).

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Example 10

Construction of recombinant adenoviral hsGC vectors

The cDNAs for hsGC $\alpha 1$ and hsGC $\beta 1$ were isolated from the original plasmid with the restriction enzyme EcoRI as 3.0 kb (hsGC $\alpha 1$) and 2.4 kb (hsGC $\beta 3$) fragments. The fragments were each inserted into the EcoRI restriction sites of the adenoviral transfer plasmid pZS2 (Figure 16), which contains an adenovirus type 5 sequence (Ad5) with a deletion in the E1 region ($\Delta E1$), followed by an expression cassette with a CMV (cytomegalovirus) promoter/enhancer and a unique XbaI restriction site. hsGC $\alpha 1$ -pZS2 and hsGC $\beta 1$ -pZS2 digested with XbaI were inserted into the XbaI site of the long arm (RR5) of Ad5 (Figure 16). The resulting recombinant adenoviral vectors Ad5CMVhsGC $\alpha 1$ and Ad5CMVhsGC $\beta 1$ are replication deficient because they lack the E1 region. To propagate the viruses, 293 cells that express E1 were infected with these viruses. Viral plaques appeared 12-24 hours after the transfection. Viruses from single plaques were purified according to a standard procedure. Plaques containing recombinant viruses (Ad5CMVhsGC $\alpha 1$ or Ad5CMVhsGC $\beta 1$) were identified by means of PCR analysis: The plaque material was freeze-thawed three times, incubated at 37°C for 30 minutes in lysis buffer (16.6 mM ammonium sulfate / 67 mM Tris-HCl pH 6.8 / 6.7 mM MgCl₂ / 5 mM 2-mercaptoethanol / 6.7 mM EDTA / 1.7 mM SDS / 50 μ g/ml proteinase K), and thereafter heat inactivated for 10 minutes at 85°C. Finally, the DNA was isolated from the lysate with a standard phenol/chloroform extraction and used for PCR analysis.

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Example 11

*Detection of cGMP formation in EA.hy926 cells after coinfection with the hsGC
adenovectors Ad5CMVhsGC α 1 and Ad5CMVhsGC β 1*

Ten 10-cm cell culture dishes with 'EA.hy926' cells were coinfectd with each of the
5 adenovectors Ad5CMVhsGC α 1 and Ad5CMVhsGC β 1 at 2×10^{10} pfu (plaque forming
units) per dish. After 72 hours, the cells were harvested by adding hypotonic lysis
buffer (25 mM triethanolamine pH 7.8 / 1 mM EDTA / 5 mM DTT / 1 μ M leupeptin /
0.5 mg/l trypsin inhibitor / 0.2 mM PMSF) and detaching with a cell scraper. The
10 homogenate was centrifuged at 500 x g for 15 minutes and the supernatant was
mixed with an equal volume of glycerol and stored at -20°C . The stimulation of hsGC
by 100 μ M SNP (sodium nitroprusside) was determined by measurement of the basal
cGMP level and the cGMP level after treatment with SNP according to the procedure
described above (see Example 5). In three samples (A, B, C), a 7-fold to 10.75-fold
elevation in the cGMP concentration compared to the basal activity was detectable
15 after SNP stimulation, while no significant elevation was measurable in the control
without adenovirus infection (Figure 17).

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